

Malaria PCR with Parasitemia Reflex, Varies

Test ID: LMALP

Explanation:

Due to low utilization and the availability of other test options, this test will be made obsolete on the effective date.

Recommended Alternative Test:

Malaria, Molecular Detection, PCR, Varies

Test ID: LCMAL

Methodology:

Real-Time Polymerase Chain Reaction (PCR)/DNA Probe Hybridization

Reference Values:

Negative

Specimen Requirements:

Both blood specimens and slides are required.

Specimen Type: Blood

Container/Tube: Lavender top (EDTA)

Specimen Volume: 4 mL

Collection Instructions:

1. Invert several times to mix blood.
2. Send specimen in original tube. Do not aliquot.

Specimen Type: Blood films

Container/Tube: Clean, grease-free slides in plastic slide container

Specimen Volume: 2 thin blood films and 2 thick blood films

Collection Instructions:

1. Ideally, blood films should be made directly from uncoagulated blood acquired via fingerstick. However, EDTA anticoagulated blood is also acceptable.
2. Prepare thin blood films as follows:
 - a. Prepare a thin film with a "feathered edge" that is no more than a single cell thick.
 - b. Allow the film to thoroughly air dry and then fix by briefly immersing in either absolute or 95% methyl alcohol.
 - c. Allow to air dry after fixation.
3. Prepare thick blood films as follows:
 - a. Place a large drop of blood (approximately the size of a dime and preferably from a fingerstick) on a slide.
 - b. Using a corner of a second slide, spread the drop in a circular motion while applying firm pressure to literally scratch the blood onto the carrier slide. This technique allows the blood to dry quickly and adhere well to the slide. Use approximately 20 circular sweeps with the second slide. The drop of blood should be about the size of a quarter when finished.
 - c. Do not fix. Air dry thoroughly (approximately 45 minutes) before placing in transport container.

Specimen Stability Information:

Specimen Type	Temperature	Time
Varies	Refrigerated (preferred)	7 days
	Ambient	7 days

CPT Code:

87798

Day(s) Performed: Monday through Sunday

Report Available: Same day/1 to 3 days

Recommended Alternative Test (if percent parasitemia is needed):

Rapid Malaria/*Babesia* Smear, Varies

Test ID: MAL

Methodology:

Giemsa Stain

Reference Values:

Negative

Specimen Requirements:

Both blood specimens and slides are required.

Specimen Type: Whole blood

Container/Tube: Lavender top (EDTA)

Specimen Volume: 1 mL

Collection Instructions:

1. Invert several times to mix blood.
2. Send whole blood specimen in original tube. **Do not aliquot.**

Specimen Type: Blood films

Slides: 2 Thin blood films and 2 thick blood films

Container/Tube: Plastic slide container

Collection Instructions:

1. Slides must be clean and grease-free.
2. Blood films should be made from fresh blood using fingerstick or drops of blood from needle following venipuncture. However, EDTA anticoagulated blood is also acceptable.
3. Prepare thin blood films as follows:
 - a. Prepare 2 thin smears with a slide prep machine. OR
 - b. Prepare a thin film with a "feathered edge" that is no more than a single cell thick.
 - c. Allow the film to thoroughly air dry and then fix by briefly immersing in either absolute or 95% methyl alcohol.
 - d. Allow to air dry after fixation.
4. Prepare thick blood films as follows:
 - a. Place a large drop of blood (approximately the size of a dime and preferably from a fingerstick) on a slide.
 - b. Using a corner of a second slide spread the drop in a circular motion while applying firm pressure to literally scratch the blood onto the carrier slide. This technique allows the blood to dry quickly and adhere well to the slide. Use approximately 20 circular sweeps with the second slide. The drop of blood should be about the size of a quarter when finished.
 - c. **Do not fix.** Air dry thoroughly (approximately 45 minutes) before placing in transport container.

Specimen Stability Information:

Specimen Type	Temperature
Varies	Refrigerated (preferred)
	Ambient

CPT Code:

87207

Day(s) Performed: Monday through Sunday

Report Available: 1 to 2 days

Questions

Contact James Conn, Laboratory Resource Coordinator at 800-533-1710.